

Mass Culturing of Stem and Bulb Nematode (*Ditylenchus dipsaci*) for use in screening and Impression Training on Carrot Discs

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Abstract— Stem and bulb nematode, *Ditylenchus dipsaci*, is a very important plant parasitic nematode, has a very wide host range and causes economic yield losses in many cultural plants in worldwide. Its races are very diverse and found in most temperate areas of the world. *D. dipsaci* is a migratory endoparasite, has the ability to enter into a dormancy stage. Genetic resistance offers one of the best control methods within the integrated pest management strategies for *D. dipsaci*. However mass rearing of stem and bulb nematode requires for the resistant studies including screening for this nematode species. The aim of study is focused on alternative rearing methods using carrot discs as a food source, culturing medium and provide a clearly outlined and visually informative guide. Carrot discs enable the rearing of high numbers of individuals of *D. dipsaci* for timely use in experiments and for screening purposes in under sterile conditions to provide a clean, same and pure source of inoculum. The carrot disc method has been shown to be suitable for stem and bulb nematode multiplication.

Keyword—Resistant, stem and bulb nematode, culture, Technique.

I. SUMMARY

Nematologists have a long time were looking to find the best methods for culture and increase population density of plant parasitic nematodes (5), but these techniques generally has not easy. Although, the three most commonly used techniques for culturing migratory endoparasitic nematodes, including alfa alfa callus (1,3,6), root expland (4), and carrot discs (5) have been published. We present in this study tried to use carrot cultur method to increase population density of nematodes. *Ditylenchus dipsaci* or stem nematode, attacks more than 1200 type of wild and cultivated plants. many agriculture plants are hosts for this species of nematode. *D. dipsaci* live mostly as a migratory endoparasite in different parts of plants like stem, leaves and flowers. In vitro rearing of *Ditylenchus dipsaci* nematodes on

carrot discs was used for grown nematodes. This technique is workable to migratory nematodes (*Ditylenchus* spp) for serial manufacture of nematode for experimental aim, direct studies of nematodes, screening trial and increase nematodes. The most advantage this method to save time that, allow a quick mass production of nematode. Plant parasitic nematode can only develop in fresh root tissue this method help *Ditylenchus dipsaci* nematode to be produce on sterilised carrot discs in petri dishes in an incubator more than in root plant under laboratory condition. Nematodes feeding two the type of plant tissue that necessary for their culture their can feed as migratory endoparasites and ectoparasites. Sedentary endoparasites nematode need individuate tissue for reproduction but it is the opposite about migratory endoparasites nematodes like *Ditylenchus dipsaci* that do not require for it and reproduce easily on undifferentiated tissue like as carrot disk and it has been indicated to be suitable tissue for their multiplication. This technique provides a basic facility for migratory endoparasites nematodes to be reared carrot discs in petri dishes inside an incubator more than in root of plant under greenhouse condition (8). Also, This study provides a protocol for the use carrot to produce and extract stem and bulb nematodes from carrot disk culture.

II. MATERIAL AND METHOD

We choose infected roots or soil with *Ditylenchus dipsaci* for extraction of the nematode to be culture. Nematodes were inoculated on the carrot disc. The suggested temperature for rapid multiplication between 19°C and 23°C depending on nematode reproduction population must be sub culture every 4-6 weeks, the development rate decreased when Temperature increased up to 30° or decrease down to 18 °C. The first to start a carrot culture, we selected clean carrots without overly thick and cracks, Washes selected carrots under distilled water, sterilise all of equipment and materials by autoclaving at 121°C for 20

minutes and sterilise the working and tools surface with 96% ethanol and hold the carrot with 96% ethanol and flame over spirit lamp repeat this for three times. Also, sterilised all of the equipment including: forceps, peeler and knife with ethanol and flaming each before used them every time. We peeled the carrot with the sterilised peeler and cut the peeled carrot into 5 mm thick section of 3-4 cm diameter and transfer the cut carrot disc into sterilised petri dishes by using the sterilised forceps. Disk must not be less than 3 cm diameter, then transfer the carrot discs into sterilised glass petri dishes (5-6 cm diameter). We stored them in the dark incubator to keep underground condition for 3-4 weeks at 19-23°C. Also, it was necessary control carrot discs best signal of healthy cultures during incubation every week. After 3-4 weeks nematodes were started to exit the carrot and when nutrient in carrots go to become empty it was indicator for harvesting the nematodes. The other words, incubation time and the initial inoculum effected how soon the nutrients become empty. For selection of nematodes we placed a concentrated of the nematode extract into a small (3-5 cm diam.) and accounted under light microscope. The number of nematode for inoculation depends how many carrots are to be required. However, it was suggested to inoculate at least 100-150 nematodes on one carrot disc and not inoculate less than 100 of them. It was recommended to inoculate at least five replicates from petri dishes. For mass multiplication we increased number of inoculated nematode on one carrot disc and selected female and male of the *Ditylenchus dipsaci* nematode and placed them in a glass petri including sterile distilled water. The aim was to deliver 50-80 nematodes per disc in a maximum of two 50 microliter smaller drops of nematode suspension transfer the nematodes. The number of nematodes in the suspension were determined the number to transfer. It was suggested use about 80-150 nematodes for mass multiplication. After that, we placed the carrot disk in an incubator for 5-6 week, after than the nematodes were harvested and stored in the fridge at 5°C, the nematode can

remain viable for up to 1-2 week. For a more completed collection, the first cut carrot disc to small pieces and placed in the petri dishes with sterile water and let the nematodes migrate freely into the petri, then extracted nematodes counted under binocular microscope and placed in the flask.

III. RESULT

The number of nematodes extracted from the 45, 60, 90 and 120 days callus culture are listed (Table 1). 2 or 3 thousands of nematodes can be extracted out of one carrot discs after 45-60 days of culture. However, the number of nematode is different use of nematode species and geographical population. The greatest number of nematodes was recovered from the 45 old day with temperature of 19 °C to 23°C, which yielded a greater than 1, 2 and 3 fold in number of nematodes that was observed from the 60, 90 and 120 days with same temperature. The lowest number of nematodes recovery was from the 120 days with 25°C to 30°C in the carrot disk that was only slightly increased from the initial inoculum levels. The number of nematodes extracted from callus after 45 days with 19°C to 23°C showed potential yields using this technique for us (Table 1). The other word, extracted of more than 3400 nematodes were observed from the 45 days with 19°C to 23°C. Therefore, temperature between 19 °C to 23°C could increase the cultivation potential of *Ditylenchus dipsaci* nematode. Also, these result indicated that, when as the number of days increased and the temperature dropped under 19°C or increased up to 23°C, the number of nematodes decreased. Because of that we observed that the stem and bulb (*ditylenchus dipsaci*) nematode was suited for culture carrot disk and the yield of eggs and increased population density of this migratory endoparasite nematode was much better by this technique than other extraction and culture methods (2, 5, 7, 8, 9). Therefore, the carrot callus method was suitable and easy to use and produce high number of *Ditylenchus dipsaci* (Fig 1).

Table.1: The effect of different days and temperature on the development of the *Ditylenchus dipsaci* nematode.

Nematode number	15-18°C	19-23°C	25-30°C
45 day-old cultur	2200	3400	1400
60 day-old culture	2000	3000	1100
90 day-old culture	1100	1400	900
120 day-old culture	600	800	500

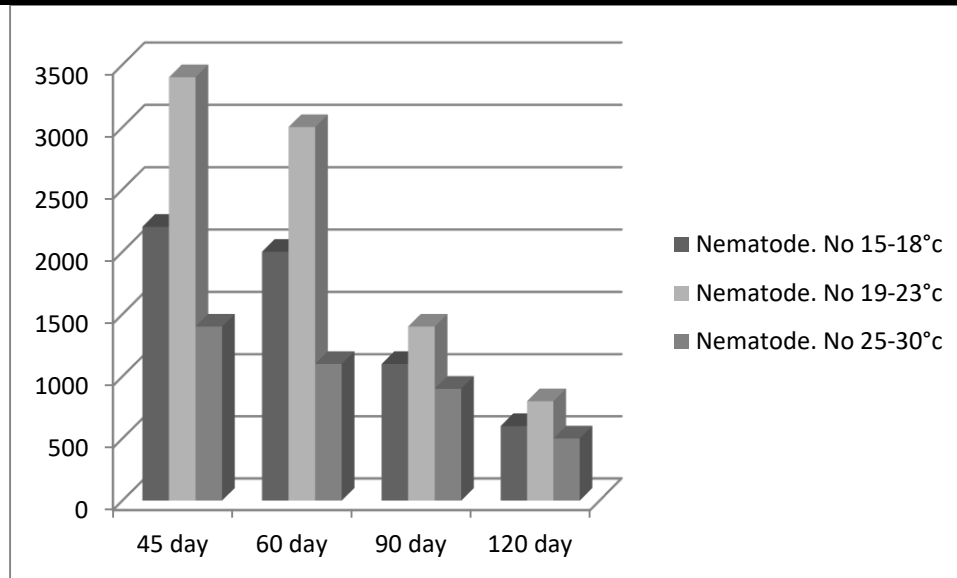


Fig.1: Mixed life stage and eggs number of *Ditylenchus dipsaci* nematode from 45,60,90 and 120 days old carrot callus culture.

IV. DISCUSSION

This protocol presented is now routinely used in our laboratory. The condition of carrots was important, work (7) has shown that carrots used in carrot disk culture must be freshly harvested from the field and greenery intact before use. Also, carrots that are badly cut must be avoided. Culture can remain available for up to 2 months and are easily generated. Once the nematode are established on the callus can be divided to small infested piece and transferred directly to make a new carrot culture. The yield of eggs and increased population density of this migratory endoparasitic nematode was much better by this technique than other extraction and culture methods (2,5,7,8,9).

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