



Development of Hatri 13DP Peanut Varieties for the Mekong Delta, Vietnam

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Received: 27 Jul 2022; Received in revised form: 20 Aug 2022; Accepted: 25 Aug 2022; Available online: 31 Aug 2022

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Abstract— The new advances are applied to production, especially by the method of marker-assisted selection (MAS) in collaboration with by improved method are made to the same breeder together many desirable genes from Yuanza 9102 / ICG 12625. From the crossed Yuanza 9102 / ICG 12625 give selected one line order named HATRI 13 DP. The time has grown HATRI 13DP (95-100 days). Height plants from 60-65 cm and compared to parents. Genotype HATRI 13DP exhibited significantly the highest pod number (78.0/plant), Hence, these traits can be selected to improve the yield potential of pole-type peanut genotypes. Weight of 100 seeds reached 42.1 g compared with their parents (Yuanza 9102 =40.9 g; ICG 12625 =36.8 g). Grain quality with red color grain protein and total fat. Productivity of HATRI 13 DP has great potential in the winter-spring season, but often for low productivity in the summer, The yield 4.2 tonnes/ha of winter-spring season and 3.5 ton/ha in Wet season. This is just like having wide adaptability, stable yield, should it be production in the Mekong delta (An Giang, Can Tho and Tra Vinh).

Keywords— *Arachis hypogaea L.*, genetic, plant breeding, marker-assisted selection, Microsatellite.

I. INTRODUCTION

Arachis hypogaea L., with the common name peanut, is an important oil, food and fodder crop grown worldwide with an annual yield of 66.3 million tons and grown on 34.1 Mha (FAOSTAT, 2021). Chromosomal peanuts (AABB, $2n = 4x = 40$), with a recent and unique polyploid origin, occurred 5 to 10 thousand years ago (Bertioli et al., 2019, 2020). This narrow genetic base and limited gene flow with its genetically diverse diploid wild relatives lead to a lack of alien strong alleles are resistant to pests in the main group of genes. However, the genetic basis underlying pod- and kernel-related traits in the peanut remained largely unknown, which hampered the improvement of peanut through marker-assisted selection. To understand the genetic basis underlying pod- and kernel-related traits in the peanut and provide more useful information for marker-assisted breeding (Weigang et al., 2016). Another limitation to recognizing yield potential is the low level of input in crop management. [World 2007]. The use of fertilizers for all crops since 2000 has averaged 11.1 kg/ha, which is not surprising enough

[Hollinger et al. 2015]. However, important factors, such as phosphorus and calcium, proved to be the leading limiting nutrients for peanut production, with only 4% and 16% of farmers using chemical fertilizers or compost, respectively, in peanut production, resulting in very low yields [World 2007]. Therefore, it is recommended to develop cultivars that tolerate deficiencies of both calcium and P in order to keep good crop yields. Soils in the Mekong Delta with an optimal pH of 6.0 to 6.5 for peanut growth (Tran et al 2021) often result in adequate availability of calcium and manganese. However, it is necessary to consider the acidic pH in the breeding program to predict increasing soil acidification in some areas of the country [Coulibaly et al. 2018] so yields are low. QTLs for productivity-related characteristics were observed and 29 QTL were identified (Yuning et al. 2017). The current QTL will contribute to a better understanding of the yield ingredients and linkage markers that will facilitate MAS breeding in peanuts. The genetic basis of the yield composition has been studied, and characteristics related to pods and seeds are highly valued in peanuts (Liu

et al. 2015; Shi et al. 2015) indicates that productivity characteristics depend on each other; Research objectives Choose to create a variety of peanuts for high yield and very ease to growing

II. MATERIAL AND METHODS

2.1. Plant materials and phenotyping

Handling Crosses of Groundnut

An F_{1-2,6-8-9} population of RIL lines was derived from a cross between Yuanza 9102/ICG 12625. The crossed seeds are grown along with their parents to identify hybrids. Plants in the F₁ generation resembling the female parent (selfed) should be removed. The plant materials (including the parents and the RILs) used in this study were originally created by our laboratory and we have all the relevant rights to the materials. The ICG 12625 variety is hardy, well-branched, stable high-yielding with a red color. HATRI 13DP crossed was started to breed in 2017 Spring-Summer 2017 season for F₁ breeding, Wet session crop 2017 F₂, Winter Spring crop 2017-2018 F₃, Spring-Summer season 2017: F₄, Summer-autumn season for self-absorption F₅, Winter-Spring crop 2018-2019 self-absorption F₆ continues to test Summer-Autumn 2018 self-absorption F₇, Winter Spring 2018-2019. Summer Autumn 2019 for Assay in Can Tho. The Seeds are harvested from F₈ generation lines, for F₉ self-absorption of the 2020 rainy season. Yields harvested in the first week of October 2020 were sown in the third week of February 2021. The experiment was conducted in farmers' fields in three provinces of An Giang (in Phu Tan); Can Tho (Binh Thuy) in which in Tra Vinh has three points (Tra Cu, ..All materials were grown in the field in accordance with the local legislation. The RIL population and its parental lines were planted in the experimental fields in Can Tho City. The field experiments followed a randomized block design with three replications according to a previous study with a few modifications (Huang et al 2015). For each plot, 10 plants from each RIL line were grown 15-cm apart within a row, and an 85-cm gap was given between RILs. The parental lines were planted after every 20 rows as controls. Standard agricultural practices were applied for field management. Each plant was harvested individually at its maturity to prevent loss from over-ripening. Only eight plants in the middle of each row were used for trait measurement. Mature seeds determined by full size pods carp color from each plant were measured for 100 seed weight, seed length, seed width and length to width ratio. The seed length and seed width were measured by using a parallel rule. The seed weight was taken on an electrical scale. The length to width ratio was calculated by dividing seed length by seed width. Number of leaves and

number of branches: calculated at the end of the harvest period, using samples of 10 plants per test. Factors constituting yield and yield: The number of pod per plant, the number of seeds per pod are determined by counting the hull and seeds of 10 plants selected from each experiment.: Grain yield: Weigh the seed mass of the plant, collect m². After harvesting, data is collected on each plot of 3m² area and weighs. The mean values of each measured trait were used for phenotypic characterization.

2.2. DNA extraction

The 100 lines F₃ were grown in pots. Maximum protection was employed to ensure healthy and disease-free seedlings. The leaves were collected 2-3 weeks after planting for DNA extraction.

Standard molecular grade chemicals and general techniques for preparing stock solutions, buffers, reagents, and equipment were followed according (Lang et al 2015)

DNA suitable for PCR analysis was prepared using a simplified procedure (McCouch *et al.*, 1988). A piece of a young rice leaf (2 cm) was collected and placed in a labeled 1.5 ml centrifuge tube in ice. The leaf was ground using a polished glass rod in a well of a spot test plate (Thomas Scientific) after adding 400 µl of extraction buffer. Grinding was done until the buffer turned green, an indication of cell breakage and release of chloroplasts and cell contents. Another 400 µl of extraction buffer was added into the well by pipetting. Around 400 µl of the lysate was transferred to the original tube of the leaf sample. The lysate was deproteinized using 400 µl of chloroform. The aqueous supernatant was transferred to a new 1.5 ml tube and DNA was precipitated using absolute ethanol. DNA was air-dried and re-suspended in 50 µl of TE buffer (Lang, 2002).

DNA quality checks used 1% agarose by melting 3 g of agarose in 300 ml of TAE buffer. The mixture was heated in a microwave for 5-6 min and then cooled to around 55-60 °C. This was then poured on a previously prepared electrophoresis box with combs. Gels were prepared and the combs removed after about 45 min. Seven microliters of DNA sample plus 3 µl of loading buffer (Tris 1 M pH = 8.0, glycerol, EDTA 0.5 M pH = 8.0, xylene cyanol 0.2%, bromophenol blue 0.2%, and distilled water) was run at 70-80 v, 60 mA for 45 min or until the loading buffer dye moved far away from the wells. The gel was then taken out and stained with ethidium bromide, after which it was observed under UV light.

Microsatellite analysis

The whole microsatellite analysis included PCR assay, polyacrylamide gel electrophoresis, and band detection and scoring.

PCR assay

Microsatellite primers were used to survey polymorphism on the samples. These were randomly selected from the 3 microsatellite primer pairs currently available for Peanut (SSR: GM 1494 (Guo và ctv 2012) :F’cttgaagaaaagtgcacg R’gaagacagaagacgaagcgtg and SNPs

Aradu_A07_1148327:F-TTGCTAATCA(G/A)TTGTTGGTTT(G/A);R-AAAGAAA(G/A)CCTTCCCCGA (Mounirou et al., 2020). enzyme *HinfI* *felowing* Lang et al 2015.)

. The PCR reaction was as follows:

Reactions were overlaid with mineral oil and processed in a programmable thermal controller set for 35 cycles of 1 min at 94 °C, 1 min at 55 °C, and 2 min at 72 °C, with a final extension at 75 °C for 5 min. After amplification, 10 µl of stop solution was added to the PCR product, which was then denatured at 94 °C for 2 min. Eight microliters of each reaction were run on polyacrylamide gel.

2.3.Data Analysis

Analysis of variance: The agro-morphological data collected were initially analyzed through analysis of variance to verify genetic variation in the traits measured. The few traits with insignificant genetic variation, based on the F-test, were not considered for further analyses.

Quality and nutritional of peanut of HATRI 13 DP and parents

Protein content was analyzed by the combustion method (Leco FP-528, 601500, Leco Corporation, St. Joseph, MI, USA) with a nitrogen conversion factor of 5.46. Moisture, oil, fiber, ash, and carbohydrate Foods 2020, 9, 942 3 of 14 contents were quantified according to the method of the Association of Official Analytical Chemists (AOAC.2000) .

Disease assessment Ten plants (third leaf on the main stem of each plant was sampled) in a single plot were scored for disease-resistant parameter evaluation based on 1-9 scale (1 = no symptoms, 2 = 1-5% leaf infection, 3 = 6-10% leaf infection, 4 = 11-20% leaf infection, 5 = 21-30 leaf infection leaf infection, 6 = 31-40% leaf infection, 7 = 41- 60% leaf infection, 8 = 61-80% leaf infection, 9 = 81-100% leaf infection) at day 100 after seed planting (Subrahmanyam et al., 1995). Disease incidence (percentage of infected plants), number and size of leaf spots (early and late blight leaf spots), %CLS leaf area of infected leaf, and defoliation were recorded for the incidence and severity of CLS in the plot.

The experiments were arranged in a completely random block style, 3 times repeated. The assay kit is performed by transplantation method (15 x 20 cm). Fertilizer : (35-60-60+150kg Ca and 40kg Mg). The seed set compares yields on a large scale according to farmers' farming techniques, applied pruning technique 20x20cm distance Agronomic indicators: Plant height: determined by cm ruler at the end of the harvest cycle (90 days) from the soil surface to the end of the main stem of 10 plants in each experiment.

III. RESULTS AND DISCUSSION

3.1. PCR-based MAS(maker assisted selection) in peanut breeding

DNA amplification by PCR - SSR method with GM 1494 marker In the Yuanza 9102/ICG 12625 population recorded P1 position corresponding to DNA of and P2: corresponding to GM 1494 shows that Yuanza 9102's DNA position is 200bp and the molecular size of ICG 12625 is 220bp. F₃ were developed from the crosses Yuanza 9102/ICG 12625 , F₃ individuals of the band pattern both parents . Line 1 gives the same size allele as the father (Select this line as HATRI 13DP). Demonstrated that on this population GM 1494 gives polymorphism.

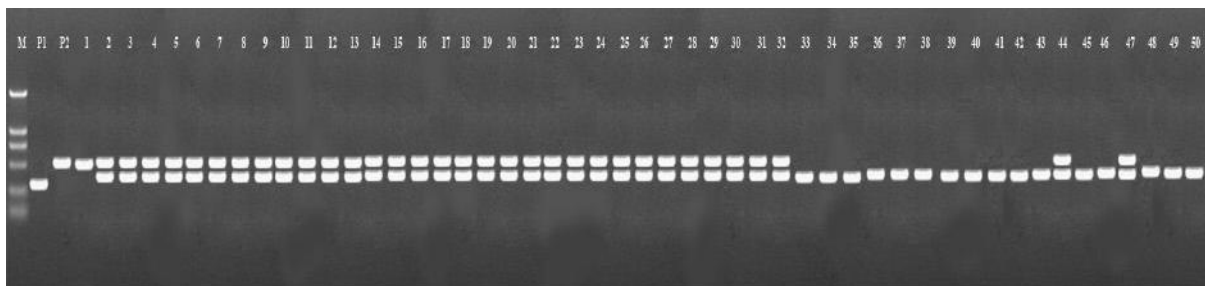


Fig.1. PCR product of the segregating population were generated by DNA amplication with primers for GM 1494 locus, on chromosome A07, positioned two the banding patterns of the parents , 220bp (ICG 12625) and 200bp (Yuanza 9102), on 3% agarose gel.

The banding patterns on the gel are the marker genotypes from which we can predict the genotype of the plants. In MAS , DNA is extracted from segregating population derived from the cross of two parents. Digest PCR products with an enzymes to detect a polymorphism between the PCR products from parents DNAs F3 position carries both parents. The tightly linked gene on chromosome A07 is marked by the molecular marker Aradu-A07-1148327. This gene is associated with the particle size group according to (Mounirou et al. (2020). Aradu-A07-1148327 molecular indicator was used on F3 populations to evaluate and select the high yield of peanuts. Markers associated with yield locus in Yuanza

9102/ICG 12625 were detected in parent Yuanza 9102/ICG 12625 for polymorphism with directive Aradu-A07-1148327. The Aradu-A07-1148327 marker is used as a marker, this marker has a size of (200-210bp) and is used as a DNA mold to establish specific primer pairs. In the recorded population with 50 plants (Fig. 3) The results showed that there were 13 lines of heterozygous form. Heterozygous carrier currents from lines (2,3,4,5,6,7,10,11,12,13,14, 16,17,18,19,20,21,22,23,24,25,26,27,28,29,30,31,32, 44,45,46) (62%) , appearing with two bands of size 200bp - 210bp corresponding to the parents.



Fig.2 : PCR product of the segregating population were generated by DNA amplication with primers for Aradu-A07-1148327 locus, on chromosome A07, positioned two the banding patterns of the parents , 200bp (ICG 12625) and 210bp (Yuanza 9102), on 3% agarose gel. Digest PCR products with HinfI enzyme Note: P1: Yuanza 9102 P2 : ICG 12625; 1-50 is a hybrid F3

3.2. Characters of growth and Morphology

Assays at HATRI Institute: In general, through provincial productivity assays in terms of yield agronomic characteristics and yield composition, the results show that

HATRI 13 is an elite peanut line, promising, short days, high yield, very good response from resistance to rust disease.

Table 1. Some agronomical features of HATRI 13 DP

No.	Parameter	Offspring HATRI 13 DP	Female parent Yuanza 9102	Male parent ICG 12625
1	Origin	Yuanza 9102 / ICG 12625	China	China
2	Growth duration (day)	85-90	80-85	80
3	Plant height (cm)	62-68	70-72	60-65
4	No. of first branch per plant	6– 6.3.0	4.0 – 5.0	5.5 - 6.0
5	Plant type	Determinant	Determinant	Determinant
6	Leaf shape	Ovate	Ovate	Ovate
7	Leaf color	Medium green	Pale green	Dark green
8	Flower color	Purple	Purple	Purple
9	Mature pot color	Yellow	Yellow	Yellow

10	Mature seed coat color	Light Yellow	Yellow	Light Yellow
11	Seed color	Red color	Dark brown	Red color
12	Diseases (score 1-9)*			
	- Rust	1	3	1
	- Black spot	1	3	1
	- Brown spot	0	1	3
13	Lodging resistance (score 1-3)	1 - 2	1	2,4-6,9
14	Rate of two seed per pod (%)	77.7	68.5	58.9
15	100-grain wgt (g)	42.1	40.9	36.8
16	Yield (ton/ha) (dry-Wet Season)	4.19-3.5	3.8-3.5	2.7-2.3

- Characters of growth and Morphology: HATRI 13 ĐP variety belongs to short duration group (85-90 days), equivalent to its parents; growing type of determinate, erect type of plant, ovate leaf shape, medium plant height (62-68 cm) and high number of first branch (2,3-3,0 branches) which is middle of parents; HATRI 13 ĐP has purple flower, big light yellow seed and brown hilum color of seed...

- Ability to resist biotic and abiotic stress: HATRI 13 ĐP has performed wide adaptability. It can be cultivated in 3 crop seasons per year (Spring, Summer and Fall-Winter); well resistant to drought (this trait was received

from its female parent, Yuanza 9102) and quite well resistant to drought, heat (this trait was received from its male parent, ICG 12625). HATRI 13 ĐP has good resistant to Rust (score 1), black spot (score 1), Brown spot (score 1-3)... ; good lodging resistance (score 1-2).

- Yield components: HATRI 13 ĐP has more number of pod per plant than that of Yuanza 9102 (27-40 pods) and high P100 seed (42.1 g) equivalent to Yuanza 9102 (40.9g) and higher than that of ICG 12625 (36.8); HATRI 13 ĐP has rate of two seed per pod higher than that of Yuanza 9102 (68.6%) and ICG 12625(58.9%).(Table 2)

Table 2. Some agronomic properties (height plants and duration) of winter-spring peanut varieties 2021

no	lines	height Plants(cm)	First flowering date (days)	Final flowering date (days)	duration(days)	Branch
1	Yuanza 9102	68.5b	38a	45a	100a	4.99c
2	ICG 12625	69.6b	37b	44b	96b	5.67b
3	HATRI 13ĐP	68.1b	37b	44b	90c	6.50a
4	MD7 (Checked)	72.0a	36c	44b	95b	5.8b

Ngày ra hoa đầu tiên

Peanuts: most varieties have an average growth time in this summer-autumn crop of 95-100 days, lasting longer than the winter-spring crop, possibly due to prolonged rains and waterlogging suitable for monoculture areas or intercropping with other crops. Flowering varieties concentrate 35-38 days after sowing, the flowers

bloom profusely during the 1st flowering Table 2 . The present study concluded that there are no significant differences on plant height with HATRI 13 ĐP with their parents , but there is significant difference in the weight of pods, % pods 3,2 and 1 seeds , number of yield provide by ANOVA test (Table 3)

Table 3: Yield and yield of components of peanut varieties dry season 2021 at Tra Cu , Tra Vinh

No	Lines	Yield (ton/ ha)	% pods 3-4seeds	% pods 2 seeds	% pods 1seeds	% pods unfilling	P100 pods (g)	P100 seeds (g)
1	Yuanza 9102	6.06a	0.00b	83.4a	10.6c	5.96c	105.8b	40.9b
2	ICG 12625	3.42c	21.5a	58.5c	12.9b	7.07b	102.8bc	36.8c

3	HATRI 13ĐP	6.19a	0.00b	77.7b	13.9a	4.48d	95.9cdef	42.1a
4	MD7 (Checked)	5.97b	0.00b	77.5b	13.9a	8.79a	92.3defg	40.2b

3.3. Multi-point assay

Assessing the genotype and environmental interactions of peanut lines on the yield of peanut varieties in the Winter Spring 2021 crop Normally a newly recognized variety must have high stability and adaptation to different environments along with high yield factors and good agronomic properties to improve variety reliability. When grown in various locations to assess their stability and adaptation, some of their agronomic characteristics and yields are likely to change. The main cause of differences in adaptability and stability between breeds is the interaction between genotype and environment. This causes a lot of difficulties in proving the superiority of a breed. The phenotype of a body is regulated through the control of the genotype and environment. The variability of the phenotype in the environment is not the same in all genotypes, the result of phenotypic variation depends on the environment.

In cases where there are multiple types of interactions, then theoretically there is only one type of interaction in which the same genotype becomes the best genotype across all environments (Chahal and Gosal, 2002). In fact, such genotypes may not be present, or can hardly develop and identify. Interference-type interactions become more realistic. Such an interaction will indicate which genotype is adapted to the environment. Non-interferometric interaction patterns affect the nature and importance of genetic variance components, on the other hand they are related to parameters such as: genetic coefficient [h²], selective efficiency [GA]. Such genotypic complexity makes improving crop yields dependent not only on the artistic ingenuity of the breeder, but also

requires a full scientific understanding of biological statistical analysis through seed assays across various sites, for rice in particular and all crops in general according to (Lang et al 2016)

Experiments evaluating adaptability and stability are usually conducted in environmental conditions other than space (place) or time (season) or both space and time. This allows us to apply the Eberhart and Russell, 1966 model to understand the interaction between genotype (G) and environment (E). Experiments evaluating gene and environmental interactions of prospective hybrid lines were conducted on a large scale. The experiment was conducted at six sites representing rice regions in the Mekong Delta: An Giang, Can Tho, Tra Vinh (three sites : Tra Cu, cau Ngang and Duyen Hai), and the experiment was conducted in two Winter Spring 2020-2021 crops. The results of rice yield assessment across 5 locations: An Giang, Can Tho, Tra Vinh, of the peanut seed/line in the Winter Spring 2021-2022 season are presented through Table 4. The results of the productivity movements showed that the F test was statistically significant at 1% in terms of the linear hypothesis of the environment, like, interacting with the environment. This allows us to use the environmental index (I_j) symbol for each place, on the interaction diagram between the genotype and the environment with the order from unfavorable to favorable as follows: Can Tho , Tra Cu, Duyen Hai, An Giang , Cau Ngang is on the I_j axis with the value in: 0.103; 0.003; 0.022; 0.017; -0.08 respectively .

Table 4: Yield (ton/ha) of peanut seed/line at 5 sites at winter-spring 2021

Lines	Can tho	Tra Cu	Duyen Hài	An Giang	Cau Ngang	Yield(ton/ha)
HATRI01ĐP	4.32	7.21	5.45	5.95	4.2	5.43
HATRI02ĐP	4.71	7.59	5.84	6.34	5.6	6.01
HATRI03ĐP	3.69	7.58	5.82	6.32	5.1	5.7
HATRI06ĐP	3.91	7.79	6.04	6.54	7.5	6.35
HATRI14ĐP	4.5	7.38	5.62	6.12	7.5	6.22
HATRI15ĐP	3.29	7.18	5.42	5.92	6.2	5.62
HATRI 13ĐP	4.94	7.81	6.06	6.56	7.8	6.63
HATRI 20ĐP	3.2	7.08	5.32	5.82	4.6	5.21
HATRI 8ĐP	3.19	7.06	5.31	5.81	5.8	5.41

MD7	2.56	7.44	5.69	5.19	6.6	5.45
Mean	3.83	7.41	5.65	6.07	6.09	5.49
IJ (Envicroments Index)	0.103	0.003	0.022	0.017	- 0.08	

In terms of rice varieties, most hybrids have an average yield higher than the control MD 7 variety (5.87 tons/ha). The difference in the yield of the varieties is significant at 5% based on a scale that evaluates productivity through multi-point analysis. The yield at Tra Cu (7.41) next Cau Ngang (6.09ton / ha), followed by An Giang (6.07). Analyzing ANOVA yields of 14 rice varieties 5 environments, the difference in the yield of varieties is very statistically significant at 1%, but the level of stability in yield, as well as adaptability manifests itself very differently, through GxE (linear) interaction is very significant. The results of the ANOVA analysis allow to consider the interaction between the breed and the environment here to be linear. In the Winter Spring 2021 season, it is clear that the yield of the lines compared to the control variety is superior such as the HATRI 13 DP line (Yield 6.63 tons / ha).

3.4. Reaction to disease

The main causes of peanut damage are rust disease is an economically important biotic stress that significantly reduces the pod and fodder yield and oil quality. It is caused by the basidiomycete fungus *Puccinia arachidis* *Sp*g. which belongs to class *Pucciniomycetes*

like other rust fungus but has fewer occurrences in teliospore form. The *P. arachidis* predominantly spreads by the repeated cycle of uredospores in the field. The disease is prevalent in most of the countries where peanut is cultivated and favored by warm and humid climatic conditions such as Tra Vinh province. Despite its economic importance, very limited work has been carried out on host-fungus interaction, fungal genetic diversity, and physiological specialization. Rust disease appears only at the end of the growth and development of peanuts. In the early stages, it is rare and the disease develops very strongly in the late stages. Peanut varieties participating in the experiment suffered from iron disease from points 1-2 points at this stage 60 after sowing, points 2-5 points 90 days after sowing.

Brown spot (*Cercospora arachidicola*), black spot (*phaeoisariopsis personala*).Brown spot and black spot disease appear earlier than rust spot disease from the start of flowering causing severe harm at the 90-day period from points 1-5 points for brown spots, 3-5 points for black spots. The MD 7 variety suffers from the most severe brown and black spot disease after 90 days .

Table 5: Resistance categories for reactions of peanut genotypes to diseases.

Lines / Disease	Rust dease			brown spot			black spot		
	30	60	90	30	60	90	30	60	90
MD7 (checked)	0	2	5	0	0	3	0	0	5
Yuanza 9102	0	1	4	0	0	2	0	0	4
ICG 12625	0	1	1	0	1	3	0	1	1
HATRI 13DP	0	0	1	0	0	0	0	0	1

3.5. Quality and nutritional of peanut of HATRI 13 DP and parents

Peanuts have a strong nutritional. They are an excellent source of plant-based protein, fiber, and many key vitamins and minerals.

- Results of analyzing seed quality indicated that in Fall-Winter crop 2021, HATRI 13 DP has good quality .The proximate composition of defatted peanuts is shown in Table 6. The largest fraction in both peanut meals was protein content, ranging from protein content of 24.5 g .Protein HATRI 13 DP was significantly higher than in

Yuanza 9102(22.3 g). Carbohydrate was the second largest fraction, found in the range of 16.13 g lower to Yuanza 9102 (17.2g) higher than ICG12625 and MD7.The total fat and crude fiber in HATRI 13 DP (43.43 and 8.4, respectively) were significantly different to those in Yuanza 9102 (39.2 and 7.8 respectively) (p < 0.05). Minerals and vitamine content can be reduced by the re-extraction process, increased contact time, or adjustment of the solid-liquid ratio. (Table 6). Minerals : postassium lower mother (Yuanza 9102)but hight father(ICG 12625) . Results of analyzing seed quality indicated that in Fall-

Winter crop 2021, HATRI 13 DP has good quality (protein content of 24.5g and lipid of 43.43 g).(Table 6)

Table 6: Nutritional value per 100 g of HATRI 13 DP and Parents

Information	Contents	HATRI 13 DP	Yuanza 9102	ICG 12625	MD7
macronutrients	protein(g)	24.5	22.3	25.2	23.2
	carbohydrate(g)	16.13	17.2	15.7	16.1
	fiber(g)	8.4	7.8	7.7	8.1
	sugars(g)	4.72	4.1	4.2	3.85
fats	Total Fat (g)	43.43	39.2	43.05	39.56
minerals	potassium(mg)	702	712	623	625
	phosphorous(mg)	356	320	378	256
	magnesium(mg)	160	152	161	153
	calcium(mg)	95	90	96	79
	sodium(mg)	17	15	17	13
	iron(mg)	4.58	5.26	5.74	3.56
	zinc(mg)	3.77	3.56	3.02	3.52
	Copper(mg)	1.356	1.421	1.253	1.461
vitamins	vitamin B-3 (niacin)(mg)	11.89	12.1	10.52	11.23
	vitamin E (alpha-tocopherol)(Mg)	8.33	7.56	8.23	8.12
	vitamin B-1 (thiamine)mg	0.64	0.55	0.62	0.6
	vitamin B-6 (pyridoxine)(mg)	0.35	0.44	0.32	0.021
	riboflavin (vitamin B-2)(mg)	0.16	0.08	0.1	0.12
	folate (vitamin B-9)(mcg)	245	215	245	241
	Vitamin C	0	0	0	0
	folate (vitamin B-9)(mcg)	245	215	245	241
	Vitamin C	0	0	0	0



Fig.3. HATRI 13DP growing at Tra Cu (Tra Vinh province)

IV. DISCUSSION

HATRI 13DP variety has significant value in height, number of branches/trees, number of seeds/trees, volume of 100 seeds, number of nodules/tree. Yield and kernel yield as well as particle size are both higher than MD 7. In addition, HATRI 13DP is a large seed cultivar, so it needs more fertilizer application than MD7. The traits of the number of seed pods per plant observed in this study confirm the findings and report significantly significant variable traits in terms of the number of seed pods per plant as reported by (Waghmode et al., 2017). the loci control the traits that determine productivity distributed across the entire genome, which also indicates a complex genetic basis of the traits that determine productivity. It is also possible to replicate QTLs with moderate phenotypic changes for seed size and weight in peanuts as well as in soybeans (Xie et al. 2014). Single-locus markers have many advantages in molecular genetics and breeding studies compared with multi-locus markers (Jin et al., 2010). The alleles of single-locus markers can be assigned to particular genomic loci in diversity analyses, preventing problems of extensive genome duplication and homology within and between different genomes caused by multi-locus markers of polyploidy (Li et al 2013). In this study, Use PCR product of the segregating population were generated by DNA amplification with primers for GM 1494 locus, on chromosome A07, positioned two the banding patterns of the parents , 220bp (ICG 12625) and 200bp (Yuanza 9102) at F3 population.

We also investigated whether the motif type, repeat length and repeat number influence the polymorphism locus SNP markers for Aradu-A07-1148327 locus, on chromosome A07 . With 4 peanut genotypes were detected showing different levels of resistance to Brown spot caused by *Cercospora arachidicola* and *P. personata*. Genotypes ranging from brown spot resistant to brown spot susceptible were observed in a field study. Differences in infected leaf area percentage among peanut genotypes provided useful information. A differential response to infection by the fungal causal agents of early and late blight existed among peanut genotypes with different inherent levels of resistance. The resistance parameters including the number of rust , brown spot and black spot , size of leaf spots, and infected leaf area percentage could differentiate genotypes into resistant and susceptible from appearance and . The results of this study indicated that genotypes HATRI 13 DP , and ICG 12625 were consistently tolerant to CLS under field conditions . Peanut is an important crop grown worldwide. Commercially it is used mainly for oil production but apart from oil, the by-products of peanut contains many other functional compounds like proteins, fibers, polyphenols, antioxidants, vitamins and minerals which can be added as a functional ingredient into many processed foods(Shalini et al., 2016) . Results of analyzing seed quality indicated that in Winter crop 2021, HATRI 13 DP has good quality (protein content of 24.5g and lipid of 43.43 g) and were

significantly different to those in Yuanza 9102 and ICG 12625 (Table 6).

V. CONSLUSIONS

Peanut variety, HATRI 13ĐP was bred using cross pollination method from Yuanza 9102/ICG 12625 , growth duration from 85-90 days (in Summer and Fall-Winter crop season), hard stem, good growth and development, wide adaptability, and can be cultivated all two seasons per year. HATRI 13ĐP has high yielding potential, high P100 seed (42.1g), with beautiful yellow seed, protein content of 24.5 g and total fat of 43.43g . Results of production testing for HATRI 13ĐP the Mekong delta provinces showed that HATRI 13ĐP has wide adaptability in many different ecological zones such as :Tra Vinh Province, An Giang Province and Can Tho City.

ACKNOWLEDGEMENTS

The authors are extremely grateful to the Tra Vinh Department of Science and Technology for providing funding to implement this topic. And High Agricultural Technology Research Institute for Mekong delta (HATRI) for all the support and facilities to conduct this experiment and the initiatives to publish the research findings.

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